

# Immune response and auto-immune diseases: gender does matter and makes the difference

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**Summary.** Women produce a more robust immune response to infection and this fact has been suggested as a tool to explain why women usually live longer than men; however, this increased immune reactivity may be responsible for the higher risk of developing autoimmune diseases (AID). Indeed, a female to male predominance occurs in AID and about 65% of all patients are women. Different factors have been implicated in underlie this striking gender difference, sex hormones being the mostly investigated. Gonadal hormones affect both the phenotype and the function of immune cells through interaction with specific receptors that are expressed in these cells. As a general rule, estrogens enhance the immune reactions, while progesterone and testosterone may exert an immunosuppressive role; however, the mechanisms involved in this complex scenario are not yet completely identified. In addition to sex hormones, genetic and environmental factors, innate and adaptive immune cells, fetal microchimerism, X chromosome inactivation and abnormalities have been proposed as key players in the development of AID and female gender bias, but their relative value is not yet fully appreciated. This review will try to critically describe the most important elements involved in the women's predominance in AID.

**Key words:** gender, immune response, autoimmune diseases, sex hormones, genetic factors, environmental factors.

## **Risposta immunitaria e malattie auto-immuni: il genere conta e fa la differenza**

**Riassunto.** Le donne producono una più intensa risposta immunitaria e ciò contribuisce a spiegare perché le donne vivano più a lungo degli uomini; tuttavia, questa aumentata reattività immunitaria predispone le donne a un aumentato rischio di sviluppare malattie autoimmuni (AID). Infatti, nelle AID, c'è una chiara predominanza del genere femminile e circa il 65% di tutti i pazienti sono donne. Tra i fattori implicati quali responsabili di questo bias di genere, gli ormoni sessuali sono sicuramente i più studiati. Gli steroidi sessuali modulano sia il fenotipo che la funzionalità delle cellule immunitarie interagendo con specifici recettori, che sono espressi nelle varie popolazioni cellulari. In linea generale, gli estrogeni potenziano le risposte immuni, mentre il progesterone e il testosterone possono esercitare un effetto di immunosoppressione; tuttavia, i meccanismi coinvolti in questo complesso scenario non sono ancora completamente identificati. Oltre agli ormoni sessuali, altri fattori (quali, ad es., alcuni determinanti genetici e ambientali, l'attività delle cellule della risposta immunitaria, il microchimerismo fetale, l'inattivazione del cromosoma X e

le anomalie cromosomiali) possono giocare un ruolo chiave nello sviluppo delle malattie autoimmuni e nel determinare il bias di genere, anche se il loro peso non è ancora completamente apprezzato. Questa review cercherà di descrivere in maniera critica i più importanti elementi che sottendono la predominanza femminile nelle malattie autoimmuni.

**Parole chiave:** genere, risposta immunitaria, malattie autoimmuni, ormoni sessuali, fattori genetici, fattori ambientali.

## Introduction

Autoimmune diseases (AID) represent an important cause of morbidity and mortality, affecting 8.5 million people in USA<sup>1,2</sup> and about 6% of the population in industrialized countries<sup>3,4</sup>.

In spite of ethnic and geographic differences in the incidence of selected AID, some groups being at higher risk for some diseases and lower risk for others<sup>4</sup>, and, despite a large variability in the age of onset, clinical setting and drug responses, most AID share a common characteristic: the prevalence of female sex.

Indeed, about 65% of all AID patients are women and this percentage is even higher in Sjogren's syndrome, systemic lupus erythematosus (SLE) and primary biliary cirrhosis<sup>4,5</sup>.

In spite of this well-known sex bias and multiple efforts to elucidate this situation, the reasons for the female predominance are still unknown. Genetic and environmental factors, innate and adaptive immune cells, sexual hormones, fetal microchimerism, X chromosome inactivation and abnormalities, have been proposed as key elements<sup>6,7</sup>, but the precise cause is still lacking.

This review will try to critically describe the most important elements involved in the women's predominance in AID.

## The immune response

As known, the immune response, a complex and tightly regulated one, is orchestrated by the immune system in order to protect the body from pathogens or other foreign damaging elements. When functioning properly, the immune system identifies a variety of threats, including viruses, bacteria and parasites, and

distinguishes them from the body's own healthy tissues. While the innate immune system represents the evolutionarily oldest defense mechanism and provides an early first line of defense against invading pathogens, the adaptive immune system allows for a stronger immune response as well as immunological memory, each pathogen being "remembered" by a signature antigen.

The components of nonspecific immune responses are monocytes, macrophages, natural killer (NK) cells, dendritic cells and granulocytes (neutrophils, eosinophils and basophils). These cells phagocytose bacteria and produce oxy-radicals (neutrophils, monocytes and macrophages), lyse infected cells (NK cells), produce cytokines to enhance nonspecific and specific immune responses. Dendritic cells, as well as monocytes and macrophages, act as antigen presenting cells (APC): they take up foreign antigens, process them and present on their surface antigen peptides for the specific immune system, mainly helper T lymphocytes.

The specific immune response comprises the humoral immune response (that is, B lymphocytes producing antibodies) and the cell-mediated immune response (that includes phagocytes, specific T lymphocytes and various cytokines).

T lymphocytes are divided into 3 distinct populations: a) cytotoxic T lymphocytes (Tc cells) that kill foreign or infected cells, b) helper T lymphocytes (Th cells) that produce cytokines and are further sub-divided into Th1 cells (producing IFN- $\gamma$  that promotes

cellular immune responses), Th2 cells (producing IL-4, IL-13 and IL-5 to help humoral immune responses) and Th17 cells (producing IL-17 that plays a key role in autoimmunity and allergen-specific responses), and c) regulatory T lymphocytes (Treg cells) that exert immunoregulation and can suppress both Th1- and Th2-mediated responses<sup>8</sup>.

The most relevant functions (as well as the major cell types involved; see also below) of innate and adaptive immunity are reported in Table 1.

Disorders of the immune system can result in immunodeficiency (when the immune system is less active than normal; chronic granulomatous disease represents a congenital, inherited immunodeficiency, whereas AIDS/HIV or immunosuppressive medications are examples of acquired immunodeficiency) or autoimmune diseases. AID represent a condition of hyperactive immune system and occur when the immune system, failing to properly distinguish between self and non-self, attacks and destroys tissues and organs of its own host.

### Cells involved in immune responses and differences between males and females

Gender differences in autoimmunity can be attributed, at least partly, to differences between male and female immune systems, women presenting stronger cellular and humoral immune reactions than men<sup>7</sup>.

**Table 1.** Major functions and relevant cell types of the innate and adaptive immune systems:

	Innate immune system	Adaptive immune system
<b>Functions</b>	<ul style="list-style-type: none"> <li>- Cell recruitment to sites of infection</li> <li>- Production of chemical mediators, including cytokines</li> <li>- Phagocytosis</li> <li>- Immune cells' activation</li> <li>- Activation of the complement cascade</li> <li>- Removal of foreign substances present in the body (by specialized cells)</li> <li>- Clearance of antibody complexes or cell debris</li> <li>- Activation of the adaptive immune system (antigen presentation)</li> <li>- Anatomical barrier (e.g., saliva, mucus, tears, sweat etc)</li> </ul>	<ul style="list-style-type: none"> <li>- Humoral immune response</li> <li>- Cell-mediated immune response</li> <li>- Recognition of specific "non-self" antigens, during the process of antigen presentation</li> <li>- Immune cells' activation</li> <li>- Generation of responses "tailored" to eliminate specific pathogens or infected cells</li> <li>- Development of immunological memory (to "remember" specific pathogens; memory B cells and memory T cells)</li> </ul>
<b>Cell types</b>	<ul style="list-style-type: none"> <li>- Phagocytes (neutrophils, monocytes, macrophages, dendritic cells)</li> <li>- Natural killer (NK) cells</li> <li>- Mast cells</li> <li>- Eosinophils</li> <li>- Basophils</li> <li>- Helper T cells</li> </ul>	<ul style="list-style-type: none"> <li>- B lymphocytes (humoral response-antibody production)</li> <li>- T lymphocytes (cell response)</li> <li>- CD8+ T cells (cytotoxic lymphocytes)</li> <li>- CD4+ T cells (helper or regulatory lymphocytes)</li> <li>- Th1 and Th2 response</li> <li>- Treg (Regulatory T) cells (control aberrant immune responses to self-antigens; autoimmune diseases)</li> <li>- Gamma delta T cells</li> </ul>

Sex differences in infectious diseases are common, but often neglected, at any age<sup>9,10</sup>, males being more susceptible than females<sup>11</sup>. This male bias has been documented for bacterial, parasitic and viral infections such as tuberculosis, leishmaniasis, leprosy, leptospirosis, HIV<sup>12,13</sup>.

Response to vaccination, too, differs, either in childhood than in adult life, between sexes: healthy females present a more robust protective antibody response to the influenza and the measles-mumps-rubella vaccines<sup>13-16</sup> and it has been also demonstrated that women could be given half dosage of the vaccine<sup>17</sup>. Thus, the enhanced female immune response to vaccination can ensure a more effective and long lasting protection but can also cause a greater prevalence of adverse effects in women<sup>14-16,18</sup>. Indeed, following monovalent 2009 pandemic influenza A (H1N1) vaccines, the female:male ratio of adverse effects was > 4:1 for healthy people aged 20-59 years<sup>18</sup>.

Sex hormones (see below) exert potent effects on immune cell subsets, estrogen and androgen receptors being present in the majority of immune cells; the "reproductive function" (including pre-puberty, puberty, pregnancy and menopause) deeply affects immune responses and AID.

As known, T lymphocytes secrete cytokines that underlie cell-mediated adaptive immunity, while B lymphocytes produce IgG and IgM antibodies.

Men and women have the same total number of lymphocytes, but males present a lower number of T cells<sup>19</sup> and post-menopausal women present less Th cells<sup>20</sup>. The overall number of B lymphocytes does not change between men and women; nevertheless, females aged > 6 years have increased IgM levels, secrete higher amounts of IL-4, IFN- $\gamma$  and IL-1, present higher CD4<sup>+</sup> T lymphocytes and higher plasma IgM levels than men<sup>7</sup> and this has been associated to the female susceptibility to AID<sup>1</sup>.

Upon antigen challenge, men's T-helper lymphocytes produce a milder "anti-inflammatory" mix of cytokines – the Th2 response, in which antibody production predominates. On the contrary, female lymphocytes tend to generate a more "pro-inflammatory" mix of cytokine, the Th1 response, in which production of cytotoxic T cells predominate, except during pregnancy. In fact, in pregnancy, women's immune system shifts towards the milder Th2 response: this may explain why some women with multiple sclerosis or rheumatoid arthritis ameliorate, especially during the third semester, while a few weeks after delivery, the disease rebounds<sup>2</sup>. Indeed, IFN- $\gamma$  production, that is the paradigm of Th1 response, is regulated by estrogens<sup>21</sup> and is secreted at higher levels after menopause and decreased over the years after reaching a plateau<sup>22</sup>. On the contrary, no difference in IL-10 production has been documented between males and females<sup>19</sup> and at different moments of the menstrual cycle<sup>23</sup>.

Higher numbers of NK cells are more often observed in women than in men<sup>24,25</sup>, their activity being modulat-

ed by estrogens in a biphasic manner: high dosage suppresses NK activity, whereas low dosage has no effect<sup>26</sup>.

Monocyte and macrophage activity is also regulated by sex steroids: estrogens stimulate TNF- $\alpha$  secretion from monocytes<sup>27</sup> whereas testosterone has no effect<sup>28</sup>. Male monocytes have been reported to produce more IL-1 $\beta$  than female ones<sup>19</sup>; this cytokine is also regulated by estrogens in a biphasic manner<sup>26</sup>. Indeed, 17 beta-estradiol modulates cytokine release through modulation of CD16 expression in human monocytes and macrophages and inhibits the release of pro-inflammatory cytokines<sup>29</sup>.

Gender differences have been observed also in polymorphonuclear leukocytes, females showing a decreased neutrophil apoptosis, as compared to males<sup>30</sup>. Moreover, chemotaxis is enhanced by progesterone, inhibited by estrogens and unaffected by testosterone<sup>31</sup>. As far as the respiratory burst is concerned, contrasting and inconclusive results have been reported<sup>26</sup>.

Gender differences have been reported also in autophagy<sup>32</sup>.

### Auto-immune diseases (AID)

Autoimmune diseases (AID) include more than 70 chronic disorders, affecting about 5% of population in the United States (with a cost of about 100 billion US dollars per year<sup>33</sup>) and presenting a large variability in terms of age of onset, targeted organs and response to therapy, but sharing a common feature: the female predominance<sup>6,11,34-36</sup>.

Indeed, as shown in Table 2, the most striking sex differences are detected in Sjogren disease, systemic lupus erythematosus (SLE), systemic sclerosis, primary biliary cirrhosis and autoimmune thyroid diseases (Graves' disease and Hashimoto's thyroiditis)<sup>4,5,34,37-39</sup>. A female to male predominance also occurs for multiple sclerosis (MS), rheumatoid arthritis (RA), dermatomyositis and myasthenia gravis<sup>37,40</sup>. On the contrary, type 1 diabetes, idiopathic pulmonary fibrosis and myocarditis are more frequent in men than in women<sup>37,41</sup> (Table2).

Also the severity of AID may vary between males and females, even if this is not so clearly defined as in the case of gender prevalence. As an example, psoriasis, SLE and disability progression in MS are more severe in males<sup>42-44</sup> and men present autoimmune hepatitis at a younger age and have high relapse rates than women<sup>45</sup>; on the contrary, Crohn's disease is more severe in girls<sup>46</sup>. Moreover, MS women have poorer survival outcomes<sup>47</sup> and relative mortality for type 1 diabetes is higher in females than in males, at least in Finland<sup>48</sup>. Increased mortality in SLE patients has been also associated with female sex<sup>49,50</sup>.

Despite the female susceptibility to AID has long been recognized, the precise cause of this bias is still

unknown and both genetic and environmental factors have been suggested as major determinants. A susceptible genetic background is necessary but does not explain by itself both AID onset and female predominance, while environmental factors act as additional players in tolerance breakdown<sup>6,51</sup>.

The most reputed mechanisms include sex hormones, fetal microchimerism, sex chromosomes and their major defects; however, none of these determinants has gathered till now enough convincing data and conflicting results are often present.

### Environmental factors

Increasing evidence supports a role for the environment in the development of AID<sup>52</sup> and at least two well-defined environmentally-associated diseases - i.e., the toxic oil syndrome that occurred after oleic anilide and 1,2-di-oleyl ester (DEPAP) addition to rapeseed oil<sup>53</sup> and the eosinophilia myalgia syndrome, occurring after ingestion of tryptophan that had been produced by an alternative manufacturing procedure<sup>54</sup> - have been described.

Infections, tobacco smoke, sun exposure, stress situations, diet and drugs have been all implicated in the development of AID<sup>3,35</sup>.

Various AID have been linked to microorganisms, e.g., *Streptococcus pyogenes* for rheumatic heart disease,

*Enterovirus* for type 1 diabetes<sup>55,56</sup>; tobacco smoke has been found to play a relevant role in some AID, as it may trigger the development of autoantibodies and act on pathogenic mechanism possibly related with an imbalance of the immune system<sup>57</sup>.

Sun exposure (i.e., ultraviolet radiation) is reported to play a role in systemic sclerosis, RA, SLE and phospholipid syndrome<sup>35</sup> and a varied sunlight exposure may occur between males and females, depending on lifestyle and/or occupation.

Cosmetics (especially hair dyes and nail polish) may also trigger primary biliary cirrhosis, an AID with a striking female predominance that affects middle-aged women, mainly<sup>58</sup>. Food intake and food composition affect immunity and auto-immunity, as vitamins and micronutrients are necessary for immune cells' development and functioning<sup>26,35</sup>. As an example, low levels of vitamin D are associated with an increased risk for MS, SLE, autoimmune thyroid diseases and others<sup>59-61</sup>.

Differences in the exposure to chemicals in the workplace between males and females are well documented and may contribute to the gender bias. As a general rule, exposure to pesticides results in anti-nuclear antibody formation<sup>62</sup>, while exposure to organic solvents is a risk factor for systemic sclerosis, primary systemic vasculitis and MS<sup>63</sup>.

### Genetic factors

Genetic polymorphisms largely contribute to AID susceptibility and may form the basis of ethnic differences in disease presentation and/or severity; as an example, in the United States, the black population presents a higher risk for SLE than whites<sup>64</sup>. Genome wide association studies (GWAS) are available for the commonest AID<sup>3,65</sup>; however, multiple genes are involved in disease susceptibility and the genetic patterns vary largely, so that most of the associations disclosed by GWAS are relatively modest<sup>3</sup>. Genetic factors may contribute to the sexual dimorphism of AID; several studies have focused on the interactions between gender and genes that affect antigen processing and presentation, lymphocyte proliferation and differentiation or encode immunoglobulins<sup>3,35</sup>.

Human leukocyte antigen (HLA) genes are located in a region that includes many genes regulating the immune response, and there is a close association between HLA genes and AID such as Graves' thyroiditis<sup>66</sup>, MS<sup>67</sup>, RA<sup>68</sup> and SLE<sup>35,69</sup>.

The majority of these associations are with HLA-DR and HLA-DQ genes, which encode for proteins that are mandatory for antigen presentation to CD4<sup>+</sup> T cells<sup>35</sup>. The association between HLA genes and AID usually presents a gender bias towards female<sup>35</sup>, with the excep-

**Table 2.** Female or male predominance in autoimmune diseases:

Autoimmune disease	Female : male ratio
Hashimoto's thyroiditis	19:1
Sjogren's syndrome	16:1
Systemic sclerosis	12:1
Primary biliary cirrhosis	10:1
Systemic lupus erythematosus	9:1
Graves' disease	7:1
Multiple sclerosis	3:1
Rheumatoid arthritis	3:1
Dermatomyositis	2:1
Myasthenia gravis	2:1
Type 1 diabetes	1:1.2
Myocarditis	1:2
Idiopathic pulmonary fibrosis	1:11



tion of SLE, where a higher HLA associated genetic risk is present in men<sup>70</sup>.

Also non-HLA genes have been associated with AID susceptibility. For instance, polymorphisms in IL-10 are associated with disease severity in RA (an AA-1087 IL-10 genotype being more frequent in females<sup>71,72</sup>) and Sjogren's syndrome<sup>73</sup>, while polymorphisms in acid phosphatase locus 1 (ACP1) and discs large homolog 5 (DLG5) have been linked to Crohn's disease<sup>74,75</sup>.

Polymorphisms in apolipoprotein E (APOE) have been related to Sjogren's syndrome (women carrying APOE epsilon 4 allele presenting an earlier onset of disease than non-carriers<sup>76</sup>), and MS<sup>77</sup>. Indeed, in this latter disease, females who have the APOE epsilon2 allele present a less severe disease<sup>78</sup>, while men carrying the APOE epsilon 4 allele experience the highest cognitive impairment<sup>79</sup>.

Due to their pivotal role in innate immunity, toll-like receptors 7 (TLR7) and 8 (TLR8), too, have been intensively investigated. As an example, following TLR7 ligation, women responded with a significantly enhanced interferon (IFN)-alpha (but not TNF-alpha) production as compared to men<sup>80</sup>.

Moreover, gender-specific association between TLR7 and TLR8 polymorphisms and TNF-alpha response after ligand stimulation were observed in measles virus and vaccine<sup>81</sup> (please, see also below).

### Sex hormones and their role in the incidence of AID

Sex hormones, as well as genes encoded on the sex chromosomes and gender-specific behavior, largely contribute to AID and influence the different immune cells by modulating their responses. The role of sex hormones and gender disparity in immunity and autoimmunity has been reviewed in a previous issue of this journal<sup>82</sup>; therefore, in the present work, I'll provide just a few examples relative to some AID.

As known, estrogens stimulate B cell production of specific antibodies in response to infection, vaccination or autoantigens<sup>41,82</sup> and may further increase the risk of AID. However, estrogen therapy in MS<sup>83</sup> and RA<sup>84</sup> may be beneficial, as well as the use of contraceptives, at least in the case of RA women < 35 years of age<sup>85</sup>. On the contrary, estrogen worsens disease severity in SLE and, in this case, blockade of ER may be beneficial<sup>86</sup>. As with the contraceptive pill, diverging results are present in the literature concerning AID<sup>35</sup>.

At high gestational levels, by inhibiting Th1 and Th17 pathways<sup>82</sup>, progesterone significantly ameliorates RA and MS<sup>87,88</sup>. Consistently, RA, which was remitted during pregnancy, usually worsens post-partum<sup>87,88</sup>. Moreover, combined estrogen and progesterone hormone replace-

ment therapy may induce lupus flares in post-menopausal women<sup>89</sup>.

As far as androgens are concerned, Klinefelter's patients (that is, males with XXY karyotype) have an increased risk to develop SLE and androgen therapy reduces immunoglobulin levels<sup>90</sup>.

One-year transdermal testosterone treatment was beneficial in MS male patients, even if it did not affect the number of lesions<sup>91</sup>, while skin patches with testosterone did not mitigate disease severity in SLE females<sup>92</sup>. In this condition, conflicting results were also reported with oral dehydroepiandrosterone, a precursor of both androgens and estrogens<sup>35</sup>. Moreover, men with RA present low testosterone levels<sup>93</sup>, men with low cortisol and androgen levels have an increased risk to develop RA<sup>94</sup> and androgen therapy in RA patients has provided some benefits<sup>90</sup>.

While mostly secreted in the anterior pituitary gland, prolactin is also produced by human lymphocytes and binds the prolactin receptor (a member of the cytokine receptor superfamily) that is located on monocytes, T and B lymphocytes<sup>35,95</sup>.

Activation of the prolactin receptor results in gene transcription, T cell proliferation and antibody secretion<sup>96</sup>. Thus, prolactin may potentiate AID, while hyperprolactinemia is often documented during different AID<sup>97</sup>. Moreover, antipsychotics-induced hyperprolactinemia is often associated with increased levels of thyroid autoantibodies<sup>98</sup>. It has been repetitively reported that bromocriptine reduces disease flares in SLE patients<sup>99-102</sup>; therefore, the issue of bromocriptine and prolactin antagonists for AID therapy warrants further investigations.

### Fetal microchimerism in autoimmunity

Microchimerism (i.e, cells' trafficking from mother to fetus and vice-versa) occurs during pregnancy and usually persists for years after delivery, fetal microchimerism being the presence of fetal cells in the maternal circulation, whereas maternal microchimerism is the persistence of maternal cells into adult life<sup>35,51,103</sup>.

A possible protective role has been proposed for fetal microchimerism, as fetal stem cells represent a potential source of cells for tissue repair, regeneration and immune suppression, but other evidences suggest that fetal microchimerism may favour neoplastic progression<sup>35,104-106</sup>. Fetal microchimerism was first evidenced in peripheral blood mononuclear cells from women with scleroderma who presented an increased level of male DNA, as compared to controls<sup>107</sup>, but this finding was not confirmed by others<sup>108</sup>.

Maternal microchimerism might result in detrimental effects, given that maternal cells are a possible source of graft vs host responses; however, it was recently shown to protect against asthma<sup>35,109</sup>.

Despite microchimerism has been observed in auto-immune thyroid diseases, type 1 diabetes, RA and other AID<sup>51,105-107,110</sup>, its role in autoimmunity and AID seems to be modest.

### Sex chromosomes, especially X chromosome

Sex determination in mammals is mediated by the Sry gene on the Y chromosome, which induces the male developmental program<sup>11</sup>. Mice with the Sry gene deleted from the Y chromosome or trans-located to an autosomal region have been used to assess the role of sex chromosomes apart from the gonadal sex<sup>11</sup>.

The X chromosome encodes about 1100 genes (that are distinct from the fewer than 100 genes on the Y chromosome)<sup>90</sup> and carries a large number of immune-related genes, including CD40L, CXCR3, OGT, FOXP3, TLR7, TLR8, IL12RG<sup>51,111-113</sup>. This is partly responsible for the female immune advantage<sup>117</sup> as, in general, women produce a more vigorous immune response to infection and this fact has been suggested as a tool to explain why women usually live longer than men<sup>2</sup>.

In females, one copy of the X chromosome is inactivated to allow equal gene expression dosage between XX females and XY males. At early development, one of the X chromosomes is silenced, resulting in a mosaic expression of either the maternal or paternal X chromosome; therefore, each X-linked gene mutation is potentially expressed in 50% cells in females but in 100% cells in males. The loss of mosaicism hypothesis states that alterations in the random X chromosome inactivation may result in autoimmunity and has been proposed to explain the female predominance in AID<sup>51,114-116</sup>.

The first support to this hypothesis came from the non-specific, polyclonal T cell activation that activated B cells presenting the same endogenous X-chromosome self antigen in females with SLE<sup>51,116-117</sup>. Indeed, the frequency of Klinefelter's syndrome (males with XXY karyotype) is 14-fold higher in men with SLE than normal men<sup>118</sup> and is comparable to the risk in females, while women with a particular X chromosome deletion (as found in the Turner's syndrome) are at lower risk for SLE<sup>119</sup>. Moreover, enhanced frequency of X monosomy has been found in women with primary biliary cirrhosis and autoimmune thyroid diseases, but not SLE<sup>120,121</sup>.

It has also been estimated that about 10% of the X chromosome escapes inactivation<sup>122</sup>: this may determine the over-expression of some gene products in females, potentially positive or negative effects depending on the gene. Over-expression and/or hypomethylation of CD40L, CXCR3 and OGT have been reported in female, but not male, SLE patients<sup>123,124</sup>. FOXP3, a gene that localizes in the short arm of the X chromosome, is essential for Treg cells and its deficiency or mutation

leads to aggressive and often fatal multi-organ AID<sup>125</sup>.

Small non-coding microRNAs (miRNAs) regulate post-transcriptional gene expression by targeting mRNAs and are emerging as new players in AID. The X chromosome (but not the Y chromosome) is highly enriched for miRNAs, whose expression can be regulated by estrogens: an altered miRNA expression has been documented in some AID, including MS, RA, SLE<sup>126,127</sup>.

In relation to autoimmunity, poor attention has been dedicated to the Y chromosome. It has been suggested to play a role in the inheritance of coronary artery disease<sup>128</sup> and has been demonstrated to undergo an age-dependent loss in some AID, including thyroid autoimmune diseases and primary biliary cirrhosis<sup>129,130</sup>.

### Conclusions

Gender differences in immunity, affecting both the innate and the adaptive immune responses, contribute to differences, between males and females, in the pathogenesis of infectious diseases, the response to vaccination and the prevalence of AID. Women have a lower burden of infections, most evident during their fertile years, but experience a higher incidence of AID. The gonadal hormones contribute to this clear gender bias, but alone are not enough. Other main players, e.g., genetic and environmental factors, sex chromosomes and their flaws, participate in such complex scenario, even if none of them has so far obtained a series of incontrovertible data, discrepant results being often reported.

Further investigation is needed to broaden our knowledge on sex and gender differences in immunity and AID; anyway, the differences so far highlighted are sufficient to suggest the need for gender-oriented therapeutic strategies in AID.

#### Key messages

- Gender differences in immune responses are well documented.
- Most auto-immune diseases share a common feature: the prevalence of the female sex.
- Immune cells express receptors for steroid sex hormones.
- Sex hormones modulate the immune responses and play a role in the onset and progression of autoimmune diseases.
- Besides sex hormones, genetic and epigenetic factors can influence the susceptibility to autoimmune diseases.

## References

- Whitacre CC, Reingold SC, O'Looney PA. A gender gap in autoimmunity. *Science* 1999; 283: 1277-8.
- McCarthy M. The "gender gap" in autoimmune disease. *Lancet* 2000; 356: 1088.
- Moroni L, Bianchi I, Lleo A. Geoepidemiology, gender and autoimmune diseases. *Autoimmunity Rev* 2012; 11: A386-A392.
- Cooper GS, Stroehla BC. The epidemiology of autoimmune diseases. *Autoimmun Rev* 2003; 2: 119-25.
- Cooper GS, Bynum MLK, Somers EC. Recent insights in the epidemiology of autoimmune diseases: improved prevalence estimates and understanding of clustering of diseases. *J Autoimmun* 2009; 33: 197-207.
- Lleo A, Battezzati PM, Selmi C, et al. Is autoimmunity a matter of sex? *Autoimmun Rev* 2008; 7: 626-30.
- Nussinovitch U, Shoenfeld Y. The role of gender and organ specific autoimmunity. *Autoimmun Rev* 2012; 11: A377-85.
- Murphy KM, Stockinger B. Effector T cell plasticity: flexibility in the face of changing circumstances. *Nat Immun* 2010; 11: 674-80.
- van Lunzen J, Altfeld M. Sex differences in infectious diseases: common but neglected. *J Infect Dis* 2014; 209 (Suppl 3): S79-S80.
- Muenchhoff M, Goulder PJR. Sex differences in pediatric infectious diseases. *J Infect Dis* 2014; 209 (Suppl 3): S120-S126.
- Markle JG, Fish EN. Sex matters in immunity. *Trends Immunol* 2014; 35: 97-104.
- Guerra-Silveira F, Abad-France, F. Sex bias in infectious disease epidemiology: pattern and processes. *PLoS One* 2013; 8: e62390.
- Klein SL, Marriott I, Fish EN. Sex-based differences in immune function and responses to vaccination. *Trans R Soc Trop Med Hyg* 2015; 109: 9-15.
- Klein SL, Jedlicka A, Pekosz A. The Xs and Y of immune responses to viral vaccines. *Lancet Infect Dis* 2010; 10: 338-49.
- Klein SL, Hodgson A, Robinson DP. Mechanisms of sex disparities in influenza pathogenesis. *J Leukoc Biol* 2012; 92: 67-73.
- Shohat T, Green MS, Nakar O, et al. Gender differences in the reactogenicity of measles-mumps-rubella vaccine. *Isr Med Assoc J* 2000; 2: 192-5.
- Engler RJ, Nelson MR, Klote MM, et al. Half- vs full-dose trivalent inactivated influenza vaccine (2004-2005): age, dose and sex effects on immune responses. *Arch Intern Med* 2008; 168: 2405-14.
- Halsey NA, Griffioen M, Dreskin SC, et al. Immediate hypersensitivity reactions following monovalent pandemic influenza A (H1N1) vaccines: Report to VAERS. *Vaccine* 2013; 31: 6107-112.
- Bouman A, Schipper M, Heineman MJ, et al. Gender difference in the non-specific and specific immune response in humans. *Am J Reprod Immunol* 2004; 52: 19-26.
- Yang JH, Liang CD, Wu MY, et al. Hormone replacement therapy reverses the decrease in natural killer cytotoxicity but does not reverse the decrease in the T-cell subpopulation of interferon-gamma production in postmenopausal women. *Fertil Steril* 2000; 74: 261-67.
- Giron-Gonzales JA, Moral FJ, Elvira J, et al. Consistent production of a higher Th1:Th2 cytokine ratio by stimulated T cells in men compared with women. *Eur J Endocrinol* 2000; 143: 31-6.
- Kamada M, Irahara M, Maegawa M, et al. Transient increase in the levels of T-helper 1 cytokines in postmenopausal women and the effects of hormone replacement therapy. *Gynecol Obstet Invest* 2001; 52: 82-8.
- Oertelt-Prigione S. Immunology and the menstrual cycle. *Autoimmun Rev* 2012; 11: A486-A492.
- Sandberg JK, Bhardwaj N, Nixon DF. Dominant effector memory characteristics, capacity for dynamic adaptive expansion, and sex bias in the innate Valpha24 NKT cell compartment. *Eur J Immunol* 2003; 33: 588-96.
- Montoya CJ, Pollard D, Martinson J, et al. Characterization of human invariant natural killer T subsets in health and disease using a novel invariant natural killer T cell-clonotypic monoclonal antibody, 6B11. *Immunology* 2007; 122: 1-14.
- Oertelt-Prigione S. The influence of sex and gender on the immune response. *Autoimmun Rev* 2012; 11: A479-A485.
- Rogers A, Eastell R. The effect of 17beta-estradiol on production on cytokine production in cultures of peripheral blood. *Bone* 2001; 29: 30-34.
- Pesma E, Moes H, Heinemann MJ, et al. The effects of testosterone on cytokine production in the specific and non-specific immune response. *Am J Reprod Immunol* 2004; 52: 237-43.
- Kramer PR, Kramer SF, Guan G. 17beta-estradiol regulates cytokine release through modulation of CD16 expression in monocytes and monocyte-derived macrophages. *Arthritis Rheum* 2004; 50: 1967-175.
- Molloy EJ, O'Neill AJ, Grantham JJ, et al. Sex-specific alterations in neutrophil apoptosis: the role of estradiol and progesterone. *Blood* 2003; 102: 2653-9.
- Miyagi M, Aoyama H, Morishita M, et al. Effects of sex hormones on chemotaxis of human peripheral polymorphonuclear leukocytes and monocytes. *J Periodontol* 1992; 63: 28-32.
- Lista P, Straface E, Brunelleschi S, et al. On the role of autophagy in human diseases: a gender perspective. *J Cell Mol Med* 2011; 15: 1443-57.
- Jacobson DL, Gange SJ, Rose NR, et al. Epidemiology and estimated population burden of selected autoimmune diseases in the United States. *Clin Immunol Immunopathol* 1997; 84: 223-43.
- Whitacre CC. Sex differences in autoimmune diseases. *Nat Immun* 2001; 2: 777-80.
- Ngo ST, Steyn FJ, McCombe PA. Gender differences in autoimmune diseases. *Frontiers in Neuroendocrinology* 2014; 35: 347-69.
- Gleicher N, Barad DH. Gender as risk factor for autoimmune diseases. *J Autoimmun* 2007; 28: 1-6.
- Hayter SM, Cook MC. Updated assessment of the prevalence, spectrum and case definition of autoimmune disease. *Autoimmun Rev* 2012; 11: 754-65.
- Brandt JE, Priori R, Valesini G, et al. Sex differences in Sjogren's syndrome: a comprehensive review of immune mechanisms. *Biol Sex Diff* 2015; 6: 19.

39. Gershwin ME, Selmi C, Worman HJ, et al. Risk factors and comorbidities in primary biliary cirrhosis: a controlled interview-based study of 1032 patients. *Hepatology* 2005; 42: 1194-202.
40. Zandman-Goddard G, Peeva E, Shoenfeld Y. Gender and autoimmunity. *Autoimmun Rev* 2007; 6: 366-72.
41. Fairweather D, Frisanco-Kiss S, Rose NR. Sex difference in autoimmune disease from a pathologic perspective. *Am J Pathol* 2008; 173: 600-9.
42. Sakai R, Matsui S, Fuhushima M, et al. Prognostic factor analysis for plaque psoriasis. *Dermatology* 2005; 211: 103-6.
43. Weinshenker BG, Rice GP, Noseworthy JH, et al. The natural history of multiple sclerosis: a geographically based study. 3. Multivariate analysis of predictive factors and models of outcome. *Brain* 1991; 114: 1045-56.
44. Crosslin KL, Wiginton KL. Sex differences in disease severity among patients with systemic lupus erythematosus. *Gen Med* 2011; 8: 365-71.
45. Al-Chalabi T, Underhill JA, Portmann BC, et al. Impact of gender on the long-term outcome and survival of patients with autoimmune hepatitis. *J Hepatol* 2008; 48: 140-7.
46. Gupta N, Bostrom AG, Kirschner BS, et al. Gender differences in presentation and course of disease in pediatric patients with Crohn disease. *Pediatrics* 2007; 120: e1418-e1425.
47. Grytten Torkidsen N, Lie SA, Aarseth JH, et al. Survival and cause of death in multiple sclerosis: results from a 50-year follow-up in Western Norway. *Multip Sci* 2008; 14: 1191-8.
48. Asao K, Sarti C, Forsen T, et al. Long term mortality in nationwide cohorts of childhood-onset type 1 diabetes in Japan and Finland. *Diabetes Care* 2003; 26: 2037-42.
49. Bernatsky S, Boivin JF, Joseph L, et al. Mortality in systemic lupus erythematosus. *Arthritis Rheum* 2006; 54: 2550-7.
50. Ruiz E, Ramalle-Gomara E, Elena A, et al. Trends in systemic lupus erythematosus mortality in Spain from 1981 to 2010. *Lupus* 2014; 23: 431-5.
51. Selmi C, Brunetta E, Raimondo MG, et al. The X chromosome and the sex ratio of auto-immunity. *Autoimmun Rev* 2012; 11: A531-A537.
52. Miller FW, Pollard KM, Parks CG, et al. Criteria for environmentally associated autoimmune diseases. *J Autoimmun* 2012; 39: 253-8.
53. Gelpi E, de la Paz M, Terracini B, et al. The Spanish toxic oil syndrome 20 years after its onset: a multidisciplinary review of scientific knowledge. *Environ Health Perspect* 2002; 110: 457-64.
54. Hertzman PA, Clauw DJ, Kaufman LD, et al. The eosinophilia-myalgia syndrome: status of 205 patients and results of treatment 2 years after onset. *Ann Intern Med* 1995; 122: 851-5.
55. Fae KC, da Silva DD, Oshiro SE, et al. Mimicry in recognition of cardiac myosin peptides by heart-intralesional T cell clones from rheumatic heart disease. *J Immunol* 2006; 176: 5662-70.
56. Richardson SJ, Willcox A, Bone AJ, et al. The prevalence of enteroviral capsid protein vp1 immunostaining in pancreatic islets in human type 1 diabetes. *Diabetologia* 2009; 52: 1143-51.
57. Perricone C, Versini M, Ben-Ami D, et al. Smoke and autoimmunity: the fire behind the disease. *Autoimmun Rev* 2016; 15: 354-74.
58. Bianchi I, Lleo A, Bernuzzi F, et al. The X-factor in primary biliary cirrhosis: monosomy X and xenobiotics. *Autoimmun Highlights* 2012; 3: 127-32.
59. Alharbi FM. Update in vitamin D and multiple sclerosis. *Neurosciences* 2015; 20: 329-35.
60. Yap KS, Morand EF. Vitamin D and systemic lupus erythematosus: continued evolution. *Int J Rheum Dis* 2015; 18: 242-9.
61. Wang X, Zynat J, Guo Y, et al. Low serum vitamin D is associated with anti-thyroid-globulin antibody in female individuals. *Int J Endocrinol* 2015; 2015: 285290.
62. Rosenberg AM, Semchuk KM, McDuffie HH, et al. Prevalence of antinuclear antibodies in a rural population. *J Toxicol Environ Health* 1999; 57: 225-36.
63. Barragan-Martinez C, Speck-Hernandez CA, Montova-Ortiz G, et al. Organic solvents as risk factor for autoimmune diseases: a systematic review and meta-analysis. *PLoS One* 2012; 7: e51506.
64. Youinou P, Pers JO, Gershwin ME, et al. Geo-epidemiology and autoimmunity. *J Autoimmun* 2010; 34: J163-167.
65. Invernizzi P, Gershwin ME. The genetics of human autoimmune diseases. *J Autoimmun* 2009; 33: 290-9.
66. Li H, Chen Q. Genetic susceptibility to Grave's diseases. *Front Biosci* 2013; 18: 1080-7.
67. Cree BA. Multiple sclerosis genetics. *Handbook Clin Neurol* 2014; 122: 193-209.
68. Jin H, Arase N, Hurayasu K, et al. Autoantibodies to IgG/HLA class II complexes are associated with rheumatoid arthritis susceptibility. *Proc Natl Acad Sci USA* 2014; 111: 3787-92.
69. Deng FY, Lei SF, Zhang YH, et al. Functional relevance for associations between genetic variants and systemic lupus erythematosus. *PLoS One* 2013; 8: e53037.
70. Hughes T, Adler A, Merrill JT, et al. Analysis of autosomal genes reveals gene-sex interactions and higher total genetic risk in men with systemic lupus erythematosus. *Ann Rheum Dis* 2012; 71: 694-9.
71. Hee CS, Gun SC, Naidu R, et al. Comparison of single nucleotide polymorphisms in the human interleukin-10 promoter between rheumatoid arthritis patients and normal subjects in Malaysia. *Mod Rheumatol* 2007; 17: 429-35.
72. Padyukov L, Hytonen AM, Smolnikova M, et al. Polymorphism in promoter region of IL-10 gene is associated with rheumatoid arthritis in women. *J Rheumatol* 2004; 31: 422-5.
73. Qin B, Wang J, Liang Y, et al. The association between TNF-alpha, IL-10 gene polymorphisms and primary Sjogren's syndrome: a meta-analysis and systemic review. *PLoS One* 2013; 8: e63401.
74. Browning BL, Annese V, Barclay ML, et al. Gender-stratified analysis of DLG5 R30Q in 4707 patients with Crohn disease and 4973 controls from 12 Caucasian cohorts. *J Med Genet* 2008; 45: 36-42.
75. Gloria-Bottini F, Bottini N, Renzetti G, et al. ACP1 and Th class of immunological diseases: evidence and interaction with gender. *Int Arch Allergy Immunol* 2007; 143: 170-6.
76. Pertovaara M, Lehtimaki T, Rontu R, et al. Presence of apolipoprotein E epsilon4 allele predisposes to early onset of primary Sjogren's syndrome. *Rheumatology* 2004; 43: 1484-7.



77. Rafiei M, Zarif Yeganeh M, Sheikholestami S, et al. Apolipoprotein E polymorphisms status in Iranian patients with multiple sclerosis. *J Neurol Sci* 2012; 320: 22-25.
78. Kantarci OH, Hebrink DD, Achenbach SJ, et al. Association of APOE polymorphisms with disease severity in MS is limited to women. *Neurology* 2004; 62: 811-4.
79. Savatteri G, Messina D, Andreoli V, et al. Gender-related effect of clinical and genetic variables on the cognitive impairment in multiple sclerosis. *J Neurol* 2004; 251: 1208-14.
80. Berghofer B, Frommer T, Haley G, et al. TLR7 ligands induce TNF-alpha production in females. *J Immunol* 2006; 177: 2088-96.
81. Clifford HD, Yerkovich ST, Khoo SK, et al. Toll-like receptor 7 and 8 polymorphisms: associations with functional effects and cellular and antibody responses to measles virus and vaccine. *Immunogenetics* 2012; 64: 219-28.
82. Ortona E, Delunardo F, Maselli A, et al. Sex hormones and gender disparity in immunity and autoimmunity. *Ital J Gender-Specific Med* 2015; 1: 45-50.
83. Sicotte NI, Liva SM, Klutch R, et al. Treatment of multiple sclerosis with the pregnancy hormone estriol. *Ann Neurol* 2002; 52: 421-8.
84. Bijlsma JW, Huber-Bruning O, Thijssen JH. Effect of estrogen treatment on clinical and laboratory manifestations of rheumatoid arthritis. *Ann Rheum Dis* 1987; 46: 777-9.
85. Spector TD, Roman E, Silman AJ. The pill, parity, and rheumatoid arthritis. *Arthritis Rheum* 1990; 33: 782-9.
86. Abdou NI, Rider V, Greenwell C, et al. Fulvestrant (Faslodex), an estrogen selective receptor downregulator, in therapy of women with systemic lupus erythematosus. Clinical, serologic, bone density, and T cell activation marker study: a double-blind placebo-controlled trial. *J Rheumatol* 2008; 35: 797.
87. Hughes GC. Progesterone and autoimmune diseases. *Autoimmun Rev* 2012; 11: A502-A514.
88. Ostensen M, Villiger PM. The remission of rheumatoid arthritis during pregnancy. *Semin Immunopathol* 2007; 29: 185-91.
89. Buyon JP, Petri MA, Kim MY, et al. The effect of estrogen and progesterone hormone replacement therapy on disease activity in systemic lupus erythematosus: a randomized trial. *Ann Intern Med* 2005; 142: 953-62.
90. Pennell LM, Galligan CL, Fish EN. Sex affects immunity. *J Autoimmun* 2012; 38: J282-J29.
91. Sicotte NI, Giesser BS, Tandon V, et al. Testosterone treatment in multiple sclerosis: a pilot study. *Arch Neurol* 2007; 64: 683-8.
92. Gordon C, Wallace DJ, Shinada S, et al. Testosterone patches in the management of patients with mild/moderate systemic lupus erythematosus. *Rheumatology* 2008; 47: 334-8.
93. Tengstrand B, Carlstrom K, Hafstrom I. Bioavailable testosterone in men with rheumatoid arthritis: high frequency of hypogonadism. *Rheumatology* 2002; 41: 285-9.
94. Masi AT, Cutolo M. Perspectives on sex hormones and the systemic rheumatic diseases. *Clin Exp Rheumatol* 1995; 13: 201-2.
95. Chavez-Rueda K, Hernandez J, Zenteno E, et al. Identification of prolactin as a novel immunomodulator on the expression of co-stimulatory molecules and cytokine secretions on T and B human lymphocytes. *Clin Immunol* 2005; 116: 182-91.
96. McMurray RW. Estrogen, prolactin and autoimmunity: actions and interactions. *Int Immunopharmacol* 2001; 1: 995-1008.
97. Orbach H, Zandman-Goddard G, Boaz M, et al. Prolactin and autoimmunity: hyperprolactinemia correlates with serositis and anemia in SLE patients. *Clin Rev Allergy Immunol* 2012; 42: 189-98.
98. Poyraz BC, Aksoy C, Balcioglu L. Increased incidence of autoimmune thyroiditis in patients with antipsychotic-induced hyperprolactinemia. *Eur Neuropsychopharmacol* 2008; 18: 667-72.
99. McMurray RW, Weidensaul D, Allen SH, et al. Efficacy of bromocriptine in an open label therapeutic trial for systemic lupus erythematosus. *J Rheumatol* 1995; 22: 2084-91.
100. Alvarez-Nemegyei J, Cobarrubias-Cobos A, Escalante-Triay E, et al. Bromocriptine in systemic lupus erythematosus: a double-blind, randomized, placebo-controlled study. *Lupus* 1998; 7: 414-9.
101. Yang XY, Liang LQ, Xu HS, et al. Efficacy of oral bromocriptine in protecting the post-partum systemic lupus erythematosus patients from disease relapse. *Zhonghua Nei Ke Za Zhi* 2003; 42: 621-4.
102. Qian Q, Liuqin L, Hao L, et al. The effects of bromocriptine on preventing postpartum flare in systemic lupus erythematosus patients from South China. *J Immunol Res* 2015; 2015: 316965.
103. Maloney S, Smith A, Furst DE, et al. Microchimerism of maternal origin persists into adult life. *J Clin Invest* 1999; 104: 41-7.
104. Nelson JL. The otherness of self: microchimerism in health and disease. *Trends Immunol* 2012; 33: 421-7.
105. Fugazzola L, Cirello V, Beck-Peccoz P. Fetal microchimerism as an explanation of disease. *Nat Rev Endocrinol* 2011; 7: 89-97.
106. Cirello V, Rizzo R, Crippa M, et al. Fetal cell microchimerism: a protective role in auto-immune thyroid diseases. *Eur J Endocrinol* 2015; 173: 111-8.
107. Nelson JL, Furst DE, Maloney S, et al. Microchimerism and Hla-compatible relationships of pregnancy in scleroderma. *Lancet* 1998; 351: 559-62.
108. Murata H, Nakauchi H, Sumida T. Microchimerism in Japanese women patients with systemic sclerosis. *Lancet* 1999; 354: 220.
109. Thompson EE, Myers RA, Du G, et al. Maternal microchimerism protects against the development of asthma. *J Allergy Clin Immunol* 2013; 132: 39-44.
110. Ye J, Vives-Pi M, Gillespie KM. Maternal microchimerism: friend or foe in type 1 diabetes? *Chimerism* 2014; 5: 21-3.
111. Libert C, Dejager L, Pinheiro I. The X chromosome in immune functions: when a chromosome makes the difference. *Nat Rev* 2010; 10: 594-604.
112. Rubtsova K, Marrack P, Rubtsov AV. Sexual dimorphism in autoimmunity. *J Clin Invest* 2015; 6: 2187-93.
113. Bianchi I, Lleo A, Gershwin ME, et al. The X chromosome and immune associated genes. *J Autoimmun* 2012; 38: J187-J192.

114. Ozcelik T. X chromosome inactivation and female predisposition to autoimmunity. *Clin Rev Allergy Immunol* 2008; 34: 348-51.
115. Kast RE. Predominance of autoimmune and rheumatic diseases in females. *J Rheumatol* 1977; 4: 288-92.
116. Stewart JJ. The female X-inactivation mosaic in systemic lupus erythematosus. *Immunol Today* 1998; 19: 352-7.
117. Takeno M, Nagafuchi H, Kaneko S, et al. Auto-reactive T cell clones from patients with systemic lupus erythematosus support polyclonal autoantibody production. *J Immunol* 1997; 158: 3529-38.
118. Scofield RH, Bruner GR, Namjou B, et al. Klinefelter's syndrome (47,XXY) in male systemic lupus erythematosus patients: support for the notion of a gene-dose effect for the X chromosome. *Arthritis Rheum* 2008; 58: 2511-7.
119. Conney CM, Bruner GR, Aberle T, et al. 46,X,del(X)(q13) Turner's syndrome women with systemic lupus erythematosus in a pedigree multiplex for SLE. *Genes Immun* 2009; 10: 478-81.
120. Invernizzi P, Miozzo M, Battezzati PM, et al. Frequency of monosomy X in women with primary biliary cirrhosis. *Lancet* 2004; 363: 533-5.
121. Invernizzi P, Miozzo M, Selmi C, et al. X chromosome monosomy: a common mechanism for autoimmune diseases. *J Immunol* 2005; 175: 575-8.
122. Lockshin MD. Non-hormonal explanations for sex discrepancy in human illness. *Ann N Y Acad Sci* 2010; 1193: 22-4.
123. Hewagama A, Gorelik G, Patel D, et al. Overexpression of X-linked genes in T cells from women with lupus. *J Autoimmun* 2013; 41: 60-71.
124. Lu Q, Wu A, Tesmer L, et al. Demethylation of CD40LG on the inactive X in T cells from women with lupus. *J Immunol* 2007; 179: 6352-8.
125. Zheng Y, Rudensky AY. FOXP3 in control of the regulatory T cell lineage. *Nat Immunol* 2007; 8: 457-62.
126. Dai R, Ahmed SA. microRNA, a new paradigm for understanding immunoregulation, inflammation, and autoimmune diseases. *Transl Res* 2011; 157: 163-79.
127. Munoz-Culla M, Irizar H, Saenz-Cuesta M, et al. SncRNA (microRNA & snoRNA) opposite expression pattern found in multiple sclerosis relapse and remission is sex dependent. *Scientific Report* 2016; 6: 20126.
128. Charchar FJ, Bloomer LD, Barnes TA, et al. Inheritance of coronary artery disease in men: an analysis of the role of Y chromosome. *Lancet* 2012; 379: 915-22.
129. Lleo A, Oertel-Prigione S, Bianchi I, et al. Y chromosome loss in male patients with primary biliary cirrhosis. *J Autoimmun* 2013; 41: 87-91.
130. Persani L, Bonomi M, Lleo A, et al. Increased loss of the Y chromosome in peripheral blood cells in male patients with autoimmune thyroiditis. *J Autoimmun* 2012; 38: 193-196.

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